IJEP 39 (7): 594-601 (2019)

Bioremediation Potential Of Macrophytes In Jakkur Wetland

- P. Sudarshan^{1,2}, M.K. Mahesh² and T.V. Ramachandra^{1,3,4}
- 1. Indian Institute of Science, Energy and Wetlands Research Group, Centre for Ecological Sciences, Bangalore-560 012
- 2. Yuvaraja's College, Department of Botany, Mysore 570 005
- 3. Indian Institute of Science, Centre for Sustainable Technologies (ASTRA), Bangalore
- 4. Indian Institute of Science, Centre for Infrastructure, Sustainable Transportation and Urban Planning (CiSTUP), Bangalore

Aquatic macrophytes are important components of wetland ecosystems as they help in the uptake of nutrients and hence help in maintaining the chemical integrity of the respective ecosystem. These plants mobilize mineral elements from the bottom sediments and provide shelter to aquatic macro invertebrates and fishes. The current study investigates the diversity, biomass, nutrient and metal uptake potential following the standard protocol. *Polygonum glabrum* and *Typha angustata* had higher biomass at both inlet and outlets. Carbon content was higher in *Typha angustata*, nitrogen and phosphorus was highest in *Spirodela polyrhiza* in the outlet. Cadmium concentration was within normal range with *Alternanthera philoxeroides* and *Pistia stratiotes* accumulating highest in inlet and outlet. *Typha angustata* in inlet had copper concentration in critical range whereas in outlet all species had normal range with *Ludwigia sp* the highest. Lead, zinc, nickel and chromium were in higher concentration in *Typha angustata*than other species in inlet. Nickel was above normal range in *Typha angustata* in inlet and in all species in outlet. In the outlet *Typha angustata* (lead), *Pistia stratiotes* (zinc and nickel) and *Alternanthera philoxeroides* (chromium) had higher concentrations. Thus the study highlighted the remediation potential of macrophytes from Jakkur lake.

KEYWORDS

Macrophytes, Bengaluru, Jakkur lake, Heavy metal

1. INTRODUCTION

Aquatic plants (macrophytes) are vascular plants growing in wetlands or on a substrate, that is where soils are flooded or saturated long enough for anaerobic conditions to develop in the root zone and these plants have evolved to adapt to an anaerobic environment [1,2]. The aquatic macrophytes occur mainly in the shallow region of lakes, ponds, pools, marshes streams and rivers, etc. Macrophytes are of considerable ecological and economic importance as they help in the uptake of nutrients and hence help in maintaining the chemical integrity of the respective ecosystem. They contribute significantly to the productivity of water bodies, mobilize mineral elements from the bottom sediments and provide shelter to aquatic macro invertebrates and fishes. Aquatic macrophytes aid in bioremediation and hence wetlands are aptly known as 'kidneys of the landscape'. They also respond to changes in water

quality and have been used as indicators of pollution and are known as 'bio-indicators'. When there is enough room for colonization and abundant availability of nutrients, macrophytes show a high growth rate. They assimilate nutrients directly into their tissues. Due to these, they were used to solve eutrophic problems of freshwater bodies and to remove pollutants [3]. Macrophytes influence water quality by taking up nutrients, releasing dissolved organic matter and increasing sedimentation by absorbing turbulent energy [4]. A considerable portion of the nutrient is stored by macrophytes and transferred to the next level (consumers) in the food chain and thus regulate the biogeochemical cycle of nutrients. However, species composition and distribution depend on environmental parameters, such as light, water temperature, substrate composition, disturbance and quality [5,6].

Macrophytes strongly influence water chemistry, acting as both nutrient sinks through uptake and also aid in moving compounds from the sediment to the water column. They improve water quality through the uptake of nutrients, trace elements and

other contaminants [7,8,9]. Aquatic macrophytes are excellent indicators of the ecological state of water bodies because they integrate environmental changes over periods of a few years and reflect the cumulative effects of successive disturbances [10]. Due to their relatively high levels of species richness, rapid growth rates and direct response to environmental changes, they are used as phyto-indicators or bioindicators of the status of water bodies [11]. Macrophytes have no mechanisms regulating uptake of nutrients, hence their impact on the environment is through a process of biochemical concentration and excretion and increased nutrient concentration in their tissues is the result of nutrient rich aquatic environment [12].

An important physiological property of aquatic vegetation, in general, is the ability to accumulate unselectively chemical elements. Taking advantage of this property, many have attempted the water purification through aquatic vegetation to remediate nutrients, heavy metals and other pollutants [13,14,15,16]. Studies reveal that the bioaccumulation of metals in a plant species also relies upon the abiotic factors, like temperature, pH and concentration of chemical elements [17]. This also helps in characterising the water body through ecological monitoring of water quality.

Metals, when discharged into the aquatic environment, undergoes physical, chemical and biological changes and binds with the particulate matter and ultimately settles in the sediment [18]. Metal accumulation in plants varies from species to species. Plants uptake metals from soil either passively through the mass flow of water into roots or direct transport through the plasma membrane of root epidermal cells [19]. Nevertheless, abilities to take up and accumulate various trace elements in the tissues depend on plant species, evident from the several studies focussing on select species [9,20,18,21,22,23,24,25]. Some species are hyper-accumulators with higher abilities to accumulate metals in their biomass. The investigations of the role of macrophytes at the landwater interface are vital as the aquatic macrophytes are involved in many ecological and environmental processes [26]. Aquatic plants are often used as bioindicators of water quality, filters of particulate matter, trapping sediments, bioremediation (removal of nutrients and heavy metals) and improvement of water quality. Phytomonitoring of chemical composition would provide insights to the uptake of nutrients during different growth phases and also the nutritional value of the plants [27,28]. The nutrient pool assessment helps in determining the

nutrients balance in the environment including uptake by primary producers [26]. The focus of the current study is to assess the phyto-diversity with the biomass and nutrient (carbon, nitrogen and phosphorus) content and to assess heavy metal (cadmium, copper, chromium, nickel, zinc and lead) uptake capability in phyto samples from Jakkur wetland, Bengaluru.

2. MATERIAL AND METHOD

2.1 Study area

Bangalore is located in the Deccan plateau, toward the south east of Karnataka state extending from 12°49′5"N to 13°8′32"N in latitude and 77°27′29" E to 77°47′2"E in longitude (Figure 1). Spatially Bangalore urban area has spatially increased from 69 km² (1901), 161 km² (1981), 221 km² (2001) to 741 km² (2006, Greater Bangalore). The undulating terrain in the region facilitated the creation of a large number of tanks in the past, providing for the traditional uses of irrigation, drinking, fishing and washing [29]. This led to Bangalore having hundreds of such water bodies through the centuries. There were 1452 water bodies in 1800 in the current spatial extent of Bangalore (741 km²). A large number of water bodies (locally called lakes or tanks) in the city had ameliorated the local climate and maintained a good water balance in the neighbourhood. The undulating topography, featured by a series of valleys radiating from a ridge, forms three major watersheds, namely the Hebbal valley, Vrishabhavathi valley and the Koramangala and Challaghatta valleys (Figure 1). These form important drainage courses for the interconnected lake system which carries storm water beyond the city limits. Bangalore, being a part of peninsular India, had the tradition of storing this water in these man-made water bodies which were used in dry periods. Today, untreated sewage is also let into these storm water streams which progressively converge into these water bodies and results in algal bloom; proliferation of exotic aquatic weeds and macrophytes; large scale fish kill due to asphyxia (zero dissolved oxygen levels) and frothing due to phosphorus enrichment [29].

The present study was conducted at Jakkur wetland in Hebbal valley and is situated at north east of Bengaluru. It spreads across 3 villages (Jakkur, Agrahara and Sampigehalli) and covers an area of 157.8 acres. It consists of 10 MLD sewage treatment plant near the inlet followed by man-made wetland through which both treated and partially treated water enters the lake (Figure 2).

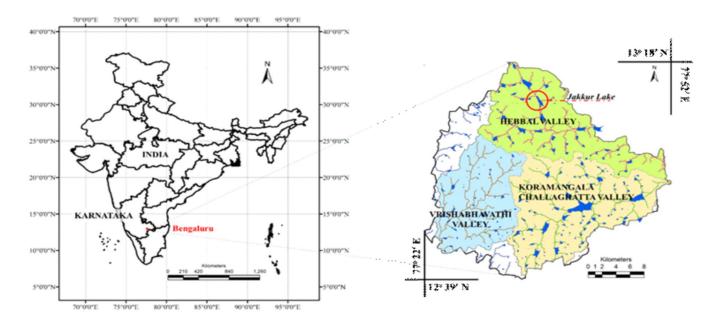


Figure 1. Study area- Jakkur lake



Figure 2. Sampling locations in the Jakkur wetlands

2.2 Bio-monitoring

Macrophytes samples were collected from inlets and outlets of Jakkur wetlands in triplicates through quadrat sampling method (0.5 m² area) every week. Plant species were identified based on morphological keys using the standard taxonomic literature and were stored in polyethene bags [30]. These samples were washed with distilled water to remove periphyton and sediments and later samples were dried at 60°C until constant weight. The dry weights of samples were noted and biomass is expressed as kg/dry weight.

2.3 Assessment of remediation potential

Dried plant samples were pulverised using mortar

and pestle, sieved (1 mm) to get fine powders and labelled properly. Later 0.5 g of samples were acid digested and analysed for six heavy metals, namely cadmium (Cd), chromium (Cr), copper (Cu), nickel (Ni), lead (Pb) and zinc (Zn) with reagent blanks and suitable standards using atomic absorption spectrophotometer (GBC Avanta version 1.31) [31]. Elemental carbon (C) and nitrogen (N) were estimated using CHN analyser. Total phosphorus was analyzed according to the standard protocol after digesting samples using HNO₃:H₂SO₄:HClO₄ [32].

3. RESULT AND DISCUSSION

3.1 Phytodiversity in Jakkur wetlands

Macrophyte distribution is related to several environmental and anthropogenic factors, such as climate, hydrology, geomorphology, nutrient availability, biological interactions and the extent of anthropogenic activities [33,34,35]. Table 1 lists 9 species of macrophytes (4 floating and 5 emergent types) belonging to 8 families (5 species from inlet and 6 species from outlet) with life forms. *Typha angustata* was the dominant species followed by *Alternanthera philoxeroides*. The percentage composition of macrophytes was given in figure 3.

3.2 Estimation of biomass

Tables 2 and 3 lists species-wise biomass and C, N and P of macrophytes samples at inlet and outlets. *Polygonum* species had higher biomass at inlet whereas *Typha angustata* had higher biomass than

Table 1. Macrophyte species with the life form and family details.

Species	Life form	Family
Typha angustata	Emergent	Typhaceae
Alternanthera philoxeroides	Emergent	Amaranthaceae
Polygonum glabrum	Emergent	Polygonaceae
Ludwigia sp	Emergent	Onagraceae
Cyperus sp	Emergent	Cyperaceae
Pistia stratiotes	Free floating	Araceae
Spirodela polyrhiza	Free floating	Lemnaceae
Lemna sp	Free floating	Araceae
Eichhornia crassipes	Free floating	Pontederiaceae

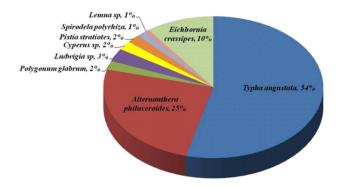


Figure 3. Species-wise distribution of macrophyte in Jakkur wetland

other species at the outlet. The biomass range was 0.15-0.412 kg/dw (*Cyperus sp. - Polygonum glabrum*) at inlet and 0.03-1.04 kg/dw (*Lemna sp - Typha angustata*) at the outlet. Among all, *Typha angustata* had higher biomass compared to others during the study period. Variations in biomass and plant zonation are due to varied growth rates among species depending on the nutrient availability and water level [36,37,38,39]. The biomass changes are low in the aquatic environment with minimal variations of water level during seasons [39].

3.3 Nutrient concentrations in macrophytes

Carbon (C), nitrogen (N) and phosphorus (P) are the most vital elements for plant morphogenesis, support a variety of cell physiological functions and their absorption and allocation are prove to be essential for all organisms [40]. These nutrients limit primary production especially in freshwater ecosystems [41]. All these species vary in their nutrient concentration and their distribution depends on nutrient availability. Plant nutrient concentration varies among sites and seasons and each species have specific ability to concentrate the nutrients.

3.3.1 Carbon: The carbon concentration ranged from 28.5-44.3% at the inlet with *Polygonum* species having higher carbon (40.7-44.3%) (Table 2). The range of carbon in the outlet was 20.5-42% with *Typha angustata* having higher carbon (37.9-42%) (Table 3).

3.3.2 Nitrogen: The concentration of nitrogen was in the range of 0.98-3.98% in the inlet (Table 2) and 1.53-4.42% (Table 3) at the outlet. Alternanthera philoxeroides (1.64-3.98%) at the inlet and Spirodela sp. (3.5-4.42%) at the oulet had higher nitrogen. The uptake potential varies widely depending on plant species and age, growing season, type of applied wastewater, environmental conditions, etc., and is related to its net productivity and the concentration in the tissues [42,43]. Nitrogen deposition increases the N:P ratio in the plants of terrestrial and freshwater ecosystems while reducing soil and water nitrogen fixation capacity and ecosystem species diversity [40].

3.3.3 Phosphorus: The concentration of phosphorus varied from 0.3-2.22% at the inlet (Table 2) and 0.95-3.52% (Table 3) at the outlet with the higher concentrations in *Typha angustata* (0.97-2.22%) at the inlet and *Spirodela* sp. (1.07-3.52%) at the outlet.

3.4 Heavy metal concentrations in macrophytes

Metal uptake by plants depends on the bioavailability of the metal in the water phase, which in turn depends on the retention time of the metal as well as the interaction with other elements and substances in water. The pH, redox potential and organic matter content in the surrounding environment of the metal bound soil or sediment will affect the tendency of the metal to exist in ionic and plant available form [44].

Table 2. Biomass and nutrient content of macrophytes at the inlet of Jakkur wetland

	Typha angustata		Alternanthera philoxeroides		Polygonum glabrum		Eichhornia crassipes		Cyperus sp	
	Range	Mean	Range	Mean	Range	Mean	Range	Mean	Range	Mean
Biomass (kg/dw)	0.184-0.412	0.29	0.16-0.276	0.2	0.22-0.45	0.33	0.18-0.29	0.256	0.15-0.16	0.157
C (%)	37.1-43.3	40.27	28.5-36.9	32.29	40.7-44.3	43.03	37-39.2	38.6	38.2-43.6	41.3
N (%)	1.52-2.84	2.08	1.64-3.98	2.85	2.58-3.12	2.85	2.15-2.5	2.2	0.98-1.46	1.25
P (%)	0.97-2.84	1.64	0.98-1.31	1.09	0.97-1.55	1.20	0.8-1.42	1.1	0.3-0.65	0.55

Table 3. Biomass and nutrient content of macrophytes at the outlet

	Typha angustata	Alternanthera philoxeroides	Ludwigia adscendens	Pistia stratiotes	Spirodela polyrhiza	Lemna sp
Biomass	0.14-1.04	0.16-0.39	0.14-0.22	0.08-0.22	0.04-0.22	0.03-0.2
(kg/dw)	0.36	0.23	0.2	0.17	0.11	0.1
C (%)	37.9-42	20.5-39.7	35.6-37.2	33.7-33.8	25.7-37.1	30.2-33.4
	40.6	35.17	36.08	33.73	32.97	31.5
N (%)	1.53-2.7	1.78-4.24	2.68-3.44	2.63-3.42	3.5-4.42	3-4.1
	1.85	3.7	3.05	3.16	4.07	3.86
P (%)	0.95-2.29	0.96-1.4	1.06-1.34	1.09-1.19	1.07-3.52	0.98-2.56
	1.31	1.06	1.2	1.12	1.12	1.6

Table 4. Heavy metal concentrations in macrophyte samples at the inlet

Metal	Range (mg	/kg)		WHO standard	Critical range in plants [47]	Normal range in plants [47]		
	Typha angustata	Alternanthera philoxeroides	Polygonum glabrum	Eichhornia crassipes	Cyperus sp			
Cd	0-0.8	0-1.8	0.2-0.8	0.1-1.8	0-0.9	0.5	5-30	0.1-2.4
Cr	5-14	0.6-10.2	8.8-12.0	3-14	0.8-10	1.3	5-30	0.03-14
Cu	0-17.2	0-2.2	0.8-1.4	0-1.5	0-1.2	40	5-30	1-5
Ni	1.4-10.4	1.4-3.2	0.2-2.4	1.5-4.1	0.1-2.1	10	10-100	0.02-5
Pb	0-9.0	0-9.8	0-5.2	0-8.5	0-4.5	2.0	30-300	0.2-20
Zn	5.2-16.4	2.8-8.6	3.2-9.0	5-14.5	1.5-8.5	60	100-400	1-400

Table 5. Heavy metal concentrations in macrophyte samples at the outlet

Species	Ranges of metal (mg/kg)						
	Cd	Cr	Cu	Ni	Pb	Zn	
Typha angustata	0-1.4	2-13.2	0-1.8	0-8.0	3.2-13.6	3.2-23.0	
Alternanthera philoxeroides	0-1.0	4.6-19.6	0-0.8	0-5.8	0-10.4	3-10.2	
Ludwigia adscendens	0.2-0.6	4-10.6	0.6-3.2	0-6.2	3.6-12.2	6.4-7.8	
Pistia stratiotes	0.2-1.6	7-9.6	1.2-2.6	6-6.2	5-8.2	7-8.8	
Spirodela polyrhiza	0-1.0	10.6-13.6	0-0.8	0-6.8	2.8-7.6	4-8.8	
Lemna sp	0-0.9	4-12.1	0-0.9	0-6.5	1.5-8.5	5-10	
Critical range in plants [47]	5-30	5-30	5-30	10-100	30-300	100-400	
Normal range in plants [47]	0.1-2.4	0.03-14	1-5	0.02-5	0.2-20	1-400	

3.4.1 Cadmium: Cadmium a highly toxic and non-essential element affects growth, metabolism and creates water stress plants [45]. Cadmium also produces oxidative stress by releasing free radicals and reactive oxygen species which cause the death of plants by damaging membrane lipids, proteins, pigments and nucleic acids [46]. Tables 4 and 5 list species-wise cadmium concentration in plants, which range from 0-1.8 mg/kg (inlet) and 0-1.6 mg/kg (outlet), which are within the normal range and lesser than values reported earlier in macrophytes of Bangalore urban lakes [47,48,49]. *Alternanthera philoxeroides* (0-1 mg/kg) and *Pistia stratiotes* (0.2-1.6 mg/kg) accumulated higher amount of cadmium.

3.4.2 Copper: Copper at low concentration is

essential for plant nutrition and is required for various enzymatic activities and are toxic at higher concentrations, leading to oxidative stress and growth inhibition [50,51]. Tables 4 and 5 list specieswise copper concen-trations of 0-17.2 mg/kg, which are higher compared to earlier studies [48]. Copper accumulation was higher in the critical range in *Typha angustata* (0-17.2 mg/kg) at the inlet. It was within the normal range in all studied species at the outlet with the highest accumulation in *Ludwigia* species (0.6-3.2 mg/kg).

3.4.3 Lead: Lead is not essential in plant organs, immobile in the soil, tends to accumulate in roots, resulting in a scarce translocation into above ground organs and is toxic [52]. Tables 4 and 5 list species-

wise lead concentration, which are within the normal range, 0-9.8 mg/kg (inlet) and 0-13.6 mg/kg (outlet). Lead accumulation was higher in *Alternanthera philoxeroides* (0-9.8 mg/kg) at inlet and *Typha angustata* (3.2-13.6 mg/kg) at outlet.

- **3.4.4 Zinc:** Zinc is a vital plant nutrient and plays a role in metabolism, plant nutrition and enzymatic activities and concentration above 500 mg/kg is phytotoxic [47]. Its toxicity in plants can lead to poor or reduced root and shoot growth as well as chlorosis of leaves [51]. Zinc was within normal range at both the sites. Tables 4 and 5 lists the values of zinc, all samples are in the range 1.5-16.4 mg/kg at inlet and 3-23 mg/kg at outlet. The concentration was higher in *Typha angustata* (5.2-16.4 mg/kg) at inlet and 3.2-23.0 mg/kg in *Typha angustata* at outlet.
- **3.4.5 Nickel:** The range of nickel was 0.1-10.4 mg/kg in inlet (Table 4) and 0-8 mg/kg in outlet (Table 5), which are above the normal range in *Typha angustata* (at inlet) and in all species at the outlet, respectively. Nickel concentration was higher in *Typha angustata* (1.4-10.4 mg/kg at the inlet and 0-8.0 mg/kg at the outlet) and *Pistia stratiotes* (6-6.2 mg/kg at the outlet), respectively. However, these concentrations were lower compared to earlier studies [48,49].
- **3.4.6 Chromium:** Tables 4 and 5 list species-wise chromium concentrations, which are in the range 0.6-19.6 mg/kg (0.6-14 mg/kg inlet; 2-19.6 mg/kg outlet). Chromium was within the normal range in inlet samples but in outlet, it was above normal range in *Alternanthera philoxeroides* species (4.6-19.6 mg/kg). The concentration was higher in *Typha angustata* (5-14 mg/kg), *Polygonum glabrum* (8.8-12.0 mg/kg) and *Eichhornia crassipes* (3-14 mg/kg) at the inlet and *Alternanthera philoxeroides* (4.6-19.6 mg/kg) at the outlet.

4. CONCLUSION

The current study provides an insight into the macrophytes diversity with the nutrient and heavy metal concentrations in samples collected from Jakkur wetland, Bangalore. Nine species of macrophytes are present with the domination of Typha angustata, Polygonum, Alternanthera philoxeroides and Typha angustata had a higher concentration of carbon, nitrogen and phosphorus, respectively at the inlet. Carbon content was higher in Typha angustata (40.6%), nitrogen (4.07%) and phosphorus (1.94%) was highest in Spirodela polyrhiza in the outlet of the lake. Cadmium was ranging from 0-1.8 mg/kg in inlet and 0-1.6 mg/kg

in outlet macrophyte samples with the Alternanthera philoxeroides and Pistia stratiotes having higher amount at both inlet and outlet. Range of copper in the studied macrophytes was 0-17.2 mg/kg with Typha angustata and Ludwigia species having higher concentrations. The concentration of lead in studied macrophyte samples were in the range 0-9.8 mg/kg (inlet) and 0-13.6 mg/kg (outlet) with Alternanthera philoxeroides accumulating the higher concentration in inlet (0-9.8 mg/kg) and Typha angustata in outlet (3.2-13.6 mg/kg). The values of zinc was in the range 1.5-16.4 mg/kg in inlet and 3-23 mg/kg in outlet samples. The concentration was higher in Typha angustata in inlet (5.2-16.4 mg/kg) and in outlet (3.2-23.0 mg/kg) samples. The range of nickel was 0.1-10.4 mg/kg in inlet and 0-8 mg/kg in outlet, respectively. Nickel concentration was higher in Typha angustata in both inlet (1.4-10.4 mg/kg) and outlet (0-8 mg/kg). The range of Chromium was 0.6-19.6 mg/kg (0.6-14 mg/kg inlet; 2-19.6 mg/kg outlet). Typha angustata (5-14 mg/kg at inlet) and Alternanthera philoxeroides (4.6-19.6 mg/kg at outlet) had a higher concentration of chromium. These results confirm bioaccumulation of heavy metals and uptake of nutrients by the macrophytes from wetlands.

ACKNOWLEDGEMENT

We acknowledge the sustained financial support for ecological research in the Western Ghats from: (i) NRDMS division, The Ministry of Science and Technology (DST), Government of India, (ii) Indian Institute of Science and (iii) ENVIS division, The Ministry of Environments, Forests and Climate Change, Government of India.

REFERENCES

- Cowardin, L.M., et al. 1979. Classification of wetlands and deepwater habitats of the United States, Washington, D.C. U.S. Department of the Interior, U.S. Fish and Wildlife Service.
- 2. Cronk, J. K. and M.S. Fennessy. 2001. Wetland plants: Biology and ecology. CRC Press.
- Sudarshan, P., M.K. Mahesh and T.V. Ramachandra. 2017. Macrophytes of Bangalore wetlands. ENVIS technical report 126. Energy and Wetlands Research Group, CES, Indian Institute of Science, Bangalore.
- 4. Schallenberg, M. and E. Waite. 2004. Survey of aquatic macrophytes in lake Waihola, summer 2002-2003. Limnology report no. 9. Department of Zoology, University of Otago.
- 5. Wetzel, R.2001. Limnology: Lake and river

- ecosystems. Academic Press, U.S.A. 1006 pp.
- 6. Jafari, M., et al. 2003. Effective environmental factors in the distribution of vegetation types in Poshtkouh rangelands of Yazd Province (Iran). J. Arid. Env., 56:627-641.
- 7. Gersberg, R.M., et al. 1986. Role of aquatic plants in wastewater treatment by artificial wetlands. Water Res., 20:363–368.
- Peverly, J.H., J.M. Surface and T. Wang. 1995. Growth and trace metal absorption by Phragmites australis in wetlands constructed for landfill leachate treatment. *Ecol. Eng.*, 5:21-35.
- Rai, U.N., S. Sinha and P. Chandra. 1999. Wastewater treatability potential of some aquatic macrophytes: Removal of heavy metals. *Ecol. Eng.*, 5(1):5-12.
- Lawniczak, A., et al. 2010. Response of emergent macrophytes to hydrological changes in a shallow lake, with special reference to nutrient cycling. Hydrobiologia. 656:243-254.
- Hellsten, S. 2001. Effects of lake water level regulation on aquatic macrophytes stands in Northern Finland and options to predict these impacts under varying conditions. *Acta Bot. Fennia.*, 171:1-47.
- Stankovic, T., et al. 2000. Concentrations of trace metals in dominant aquatic plants of lake Provala (Vojvodina, Yugoslavia). Biologia Plant. 43:583-585.
- DeBusk, T.A., F.E. Dierberg and K.R. Reddy. 2001. The use of macrophyte based system for phosphorus removal: An overview of 25 years of research and operational results in Florida. Water Sci. Tech., 44(11-12):39-46.
- Nirmal Kumar, J.I., et al. 2008. Macrophytes in phytoremediation of heavy metal contaminated water and sediments in Pariyej community reserve, Gujarat. Turk. J. Fish. Aquat. Sci., 8:193-200.
- Srivastava, S., et al. 2014. Effect of combinations of aquatic plants (Hydrilla, Ceratophyllum, Eichhornia, Lemna and Wolffia) on arsenic removal in field conditions. Ecol. Eng., 73:97-301.
- Pawan, K.G., N. Kumar and M. Kumar. 2015. Phytoremediation of wastewater through aquatic plants for the change detection analysis in the chemical properties within the district Dhanbad, Jharkhand. *Int. J. Res. Eng. Tech.*, 4(2):243-252.
- 17. Matagi, S.V., D. Swai and R.Mugabe. 1998. A review of heavy metal removal mechanisms in

- wetlands. Afr. J. Trop. Hydrobiol. Fish., 8:23-35.
- Rai, P.K. 2009. Heavy metals in water, sediments and wetland plants in an aquatic ecosystem of tropical industrial region, India. *Env. Monit. Assess.*, 158(1-4):433-457.
- 19. Yoon, J., et al. 2006. Accumulation of Pb, Cu, and Zn in native plants growing on a contaminated Florida site. Sci. Total. Env., 368(2-3):456-464.
- 20. Lee, C.R., T.C. Sturgis and M.C. Landin. 1981. Heavy metal uptake by marsh plants in hydroponic solution cultures. *J. Plant. Nutrition.*, 3(1-4):139-151.
- Zayed, A., S. Gowthaman and N. Terry. 1998. Phytoaccumulation of trace elements by wetland plants: I. Duckweed. *J. Env. Qual.*, 27(3):715-721.
- 22. Qian, J.H., et al. 1999. Phytoaccumulation of trace elements by wetland plants: III. Uptake and accumulation of ten trace elements by twelve plant species. J. Env. Qual., 28(5):1448-1455.
- 23. Zhu, Y.L., *et al.* 1999. Phytoremediation of trace elements by wetland plants: II. Water hyacinth. *J. Env. Qual.*, 28(1):339-344.
- 24. Wang, Q., Y. Cui and Y. Dong. 2002. Phytoremediation of polluted waters: Potentials and prospects of wetland plants. *Acta. Biotech.*, 22(1-2):199-208.
- 25. Kamal, M., et al. 2004. Phytoaccumulation of heavy metals by aquatic plants. Env. Int., 29:1029-1039.
- 26. Mara, L.R.C. and H. Raoul. 2010. Phosphorus, nitrogen and carbon contents of macrophytes in lakes lateral to a tropical river (Paranapanema river, Sao Paulo, Brazil). *Acta Limnol. Bras.*, 22(2):122-132.
- 27. Barbieri, R. and F.A. Esteves. 1991. The chemical composition of some aquatic macrophyte species and implications for the metabolism of a tropical lacustrine ecosystem Lobo reservoir, Sao Paulo, Brazil. *Hydrobiologia*., 213:133-140.
- 28. Gerloff, G.C. and P.H. Krombholz. 1996. Tissue analyses as a measure of nutrient availability for the growth of angiosperm aquatic plants. *Limnol. Oceanogr.*, 11:529-537.
- 29. Ramachandra, T.V., et al. 2017. Rejuvenation blueprint for lakes in Vrishabhavathi valley. ENVIS technical report 122. Environmental Information System, CES, Indian Institute of

- Science, Bangalore.
- 30. Cook, C.D.K. 1996. Aquatic and wetland plants of India. Oxford University Press, New Delhi.
- 31. APHA. 1995. Standard methods for the examination of water and wastewater, (19th edn). American Public Health Association, Washington.
- Tandon, H.L.S. 1993. Methods of analysis of soils, plants, waters and fertilizers. Fertilizer Development and Consultation Organization, New Delhi.
- 33. LacouL, P. and B. Freedman. 2006. Environmental influences on aquatic plants in freshwater ecosystems. *Env. Rev.*, 136:89-136.
- 34. Feldmann, T. and P. Noges. 2007. Factors controlling macrophyte distribution in large shallow lake Vortsjarv. *Aquat. Bot.*, 87(1):15-21.
- 35. Hrivnak, R., H. Otaheova and D. Gomory. 2009. Seasonal dynamics of macrophyte abundance in two regulated streams. *Cent. Eur. J. Biol.*, 4(2):241-249.
- 36. Pompeo, M.L.M., R. Henry and C.V. Moschini. 2001. The water level influence on biomass of Echinochloa polystachya (poaceae) in the Jurumirim reservoir (Sao Paulo, Brazil). *Rev. Brasil. Biol.*, 61(1):19-26.
- 37. Blom, C. W. P. M., *et al.* 1990. Adaptations to flooding in plants from river areas. *Aquat. Bot.*, 38: 29-47.
- 38. Junk, W. J. and M.T.F. Piedade. 1993. Biomass and primary-production of herbaceous plant communities in the Amazon floodplain. *Hydrobiologia.*, 263:155-162.
- 39. Camargo, A.F.M. and F.A. Esteves. 1996. Influence of water level variation on biomass and chemical composition of the aquatic macrophyte Eichhornia azurea (Kunth) in an oxbow lake of the Rio Mogi-Guaçu (Sao Paulo, Brazil). *Archiv. Hydrobiol.*, 135:423-432.
- 40. Sardans, J., A. Rivas Ubach and J. Penuelas. 2012. The elemental stoichiometry of aquatic and terrestrial ecosystems and its relationships with organismic lifestyle and ecosystem structure and function; A review and

- perspectives. Biogeochemistry., 111:1-39.
- 41. Maathuis, F.J. 2009. Physiological functions of mineral macronutrients. *Curr. Opin. Plant. Biol.*, 12:250-258.
- 42. Vymazal, J. 2007. Removal of nutrients in various types of constructed wetlands. *Sci. Total. Env.*, 380:48-65.
- 43. Maurizio B. and S. Michela. 2012. Effects of five macrophytes on nitrogen remediation and mass balance in wetland mesocosms. *Ecol. Eng.*, 46:34-42.
- 44. Jackson, L.J. and J. Kalff. 1993. Patterns in metal content of submerged aquatic macrophytes; the role of plant growth form. *Fresh. Biol.*, 29:351-359.
- 45. Divan, A.M., et al. 2009. Using wild plant species as indicators for the accumulation of emissions from a thermal power plant, Cadiota, South Brazil. *Ecol. Indic.*, 9:1156-1162.
- 46. Foyer, C.H., M. Lelandais and K.J. Kunert. 1994. Photooxidative stress in plants. *Physiol. Plant.*, 92:696-717.
- 47. Maiti, S.K. 2003. Handbook of methods in environmental studies. In Air, noise, soil and overburden analysis (vol 2). ABD Publishers.
- 48. Jumbe, A.S. and N. Nandini. 2012. Heavy metals accumulation in macrophytes in the lakes of Bangalore Urban. *Ecoscan.*, 6:41-45.
- 49. Ramachandra, T.V., *et al.* 2018. Spatial patterns of heavy metal accumulation in sediments and macrophytes of Bellandur wetland, Bangalore. *J. Env. Manage.*, 206:1204-1210.
- 50. Bonanno, G. 2013. Comparative performance of trace element bioaccumulation and biomonitoring in the plant species Typha domingensis, Phragmites australis and Arundo donax. *Ecotoxicol. Env. Saf.*, 97:124-130.
- 51. Gill, M. 2014. Heavy metal stress in plants: A review. *Int. J. Adv. Res.*, 2:1043-1055.
- 52. Bonanno, G. and R. Lo Giudice. 2010. Heavy metal bioaccumulation by the organs of Phragmites australis (common reed) and their potential use as contamination indicators. *Ecol. Indic.*, 10:639-645.